



## INTERNATIONAL APPLICATION PUBLISHED UNDER THE PATENT COOPERATION TREATY (PCT)

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<b>(21) International Application Number:</b> PCT/FI90/00015 <b>(22) International Filing Date:</b> 15 January 1990 (15.01.90) <b>(30) Priority data:</b> 297791 17 January 1989 (17.01.89) US <b>(71) Applicant:</b> CULTOR LTD. [FI/FI]; Kyllikinportti 2, SF-00240 Helsinki (FI). <b>(72) Inventors:</b> HEIKKILÄ, Heikki ; Allonkohina 4 C 27, SF-02320 Espoo (FI). NURMI, Juha ; Vasarasepantie 4 D 6, SF-10330 Pinjainen (FI). RAHKILA, Leena ; Koivuvuoriantie 30 A 6, SF-02130 Espoo (FI). TÖYRYLÄ, Marja ; Harakantie 12, SF-02440 Kirkkonummi (FI).		<b>(74) Agent:</b> OY KOLSTER AB; Stora Robertsgatan 23, P.O. Box 148, SF-00121 Helsinki (FI).  <b>(81) Designated States:</b> AT (European patent), AU, BE (European patent), BR, CA, CH (European patent), DE (European patent), DK (European patent), ES (European patent), FI, FR (European patent), GB (European patent), IT (European patent), JP, LU (European patent), NL (European patent), NO, SE (European patent), SU.  <b>Published</b> <i>With international search report.</i>
<b>(54) Title:</b> METHOD FOR THE PRODUCTION OF XYLITOL FROM MIXTURES CONTAINING XYLOSE  <b>(57) Abstract</b>  A method for the production of xylitol from an aqueous xylose solution which comprises the steps of fermenting said aqueous xylose solution with a yeast strain capable of converting free xylose to xylitol, and free hexoses present in said solution to ethanol, for a period of time sufficient to produce a fermented solution containing xylitol; and separating one or several xylitol-rich fraction by chromatographic separation from said fermenting solution.		

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Method for the production of xylitol from mixtures containing xylose.

This invention relates to a method for the production of xylitol from a xylose and/or xylan-containing material, and in particular, from biomass hemicellulose hydrolysates. A specific aspect of this invention relates to a method for the production of xylitol by fermentation of biomass hemicellulose hydrolysates with a yeast strain capable of converting free xylose to xylitol and enrichment of the xylitol concentrations by chromatographic separation for crystallization.

The pentahydric alcohol xylitol is the sugar alcohol derived from the reduction of xylose ( $C_5H_{10}O_5$ ). Xylitol is a naturally occurring, five-carbon sugar alcohol which has the same sweetness and caloric content of sugar (4 kilocalories per gram). Xylitol is found in small amounts in many fruits and vegetables and is produced in the human body during normal metabolism. Xylitol has certain known metabolic, dental and technical characteristics which make it an attractive sugar substitute in various contexts.

Xylitol is metabolized independently of insulin, so it can be safely consumed by non-insulin dependent diabetics. Further, xylitol has been shown to delay gastric emptying and to possibly suppress food intake which means it may have an important role in weight reducing diets.

Xylitol is also a non-cariogenic, and possibly even a cariostatic substance. In the mouth, sucrose and other carbohydrates are fermented by *Streptococcus mutans* and other bacteria, generating acid which lowers the pH, demineralizes tooth enamel and leads to dental caries. *S. mutans* and other bacteria do not, however, ferment xylitol thereby generating the acid by-products of fermentation which contribute to tooth decay. Studies have also produced data which suggests that xylitol may even actively

suppress the formation of new caries and may even "reverse" existing lesions by inducing remineralization.

From a taste perspective, xylitol does not typically manifest an unpleasant aftertaste like other sugar substitutes and, because of the high energy required to dissolve one gram of xylitol, it produces a pleasant "cooling" effect in the mouth.

Despite xylitol's advantages, the utilization of xylitol on a commercial scale has been limited by its relatively high cost, due to the difficulty of its production on a commercial scale. Xylitol is generally prepared from xylan-containing material, particularly hydrolysates of hemicelluloses. Hemicelluloses are a large group of well characterized polysaccharides found in the primary and secondary cell walls of all land and freshwater plants. Hemicelluloses are made up of sugar residues, among others D-xylose and including D-mannose, D-glucose, D-galactose and L-arabinose.

In prior art methods, xylitol has been prepared from xylan-containing material by hydrolyzing the material to produce a mixture of monosaccharides, including xylose. The xylose is converted to xylitol, generally in the presence of a nickel catalyst such as Raney-nickel. The prior art reveals a number of methods for the production of xylose and/or xylitol from xylan-containing material. Included in such prior art methods are U.S. Patent No. 3,784,408 (Jaffe et al.), U.S. Patent No. 4,066,711 (Melaja et al.), U.S. Patent No. 4,075,405 (Melaja et al.), U.S. Patent No. 4,008,285 (Melaja et al.) and U.S. Patent No. 3,586,537 (Steiner et al.).

These prior art methods are, however, complicated multistep processes which are relatively expensive and inefficient. The prior art recognizes one of the principal problems in this context is the efficient and complete separation of xylose and/or xylitol from polyols and other

by-products of hydrolysis in order to obtain xylitol of sufficient purity. In order to address this fundamental concern, multistep separation techniques, including mechanical filtration and chromatographic separation are generally required. In addition, the art teaches the use of other purification methods, such as the use of acids to precipitate lignins which generally increase the time and expense of xylitol production on a commercial scale.

It is known that certain yeast possess the enzyme xylose reductase which catalyzes the reduction of D-xylose to xylitol as the first step in D-xylose metabolism. Studies, on an experimental scale, have utilized yeast cells capable of fermenting D-xylose or cell-free extracts containing xylose reductase to produce xylitol from D-xylose rich starting material. Gong, et al., Quantitative Production of Xylitol From D-Xylose By a High Xylitol Producing Yeast Mutant *Candida Tropicalis* HXP2, *Biotechnology Letters*, Vol. 3, No. 3, 125-130 (1981); Kitpreechavanich, V. et al.: Conversion of D-Xylose Into Xylitol By Xylose Reductase From *Candida Pelliculosa* Coupled With the Oxidoreductase System of Methanogen Strain HU, *Biotechnology Letters*, Vol. 10, 651-656 (1984); McCracken and Gong, Fermentation of Cellulose and Hemicellulose Carbohydrates by Thermotolerant Yeasts, *Biotechnology and Bioengineering Symp.* No. 12, pp. 91-102 (John Wiley & Sons 1982). Although yeast strains exist which are capable of producing high yields of xylitol from the fermentation of D-xylose, a complete process for producing xylitol from, for example, biomass hemicellulose hydrolysates which contain xylose in addition to hexoses and other impurities on a commercial scale has not been disclosed by the prior art.

The present invention, however, discloses a simple and efficient method of producing pure xylitol from xylose-containing starting material which utilizes yeast strains capable of converting xylose to xylitol and most

hexoses present to ethanol; such fermentation produces a xylitol rich solution from which xylitol can be simply and efficiently purified without resort to any extensive and expensive separation expedients. Generally, the xylitol  
5 can be purified in one step by chromatographic separation and subsequently crystallized to form pure xylitol. Small amounts of ethanol are easily removed by evaporation or similar expedients, thereby avoiding the need for extensive techniques to separate the xylitol from hexitols and  
10 other sugars generated by hydrolysis and conventional hydrogenation and which are present in the xylitol rich solution.

The present invention contemplates a method for the production of substantially pure xylitol from an aqueous  
15 xylose solution which may also contain hexoses such as glucose as well as other impurities. The invention contemplates fermenting said solution using a yeast strain capable of converting substantially all of said free xylose to xylitol and most of said free hexoses to ethanol.  
20 The fermented product is purified by removing yeast cells from the solution by filtration, centrifugation or other suitable expedients and removing ethanol by evaporation or distillation. Chromatographic separation yields a xylitol rich fraction or fractions from which pure xylitol can be  
25 crystallized.

In some cases, pretreatment of the aqueous xylose solution is utilized. Such pretreatment may include post-hydrolysis and/or separation steps in order to remove components which may be toxic and/or harmful to the yeast  
30 used to convert the xylose to xylitol or other impurities which may adversely affect the subsequent fermentation and separation steps. Such pretreatment steps can include chromatographic separation techniques.

The present invention utilizes yeasts capable of  
35 reducing xylose to xylitol and hexoses to ethanol. Such

yeasts include, but are not limited to, yeasts of the genus *Candida*, *Pichia* and *Pachysolen* and *Debaryomyces*. Of these genera, *Candida* and *Debaryomyces* are preferred with *Candida tropicalis* and *Debaryomyces hansenii* being particularly preferred. One good example is the *Candida tropicalis* strain deposited in the American Type Culture Collection and assigned Accession Number 9968.

Detailed Description of the Preferred Embodiment

A. General

1. Starting Materials

The starting materials for use with the methods of the present invention include almost any xylan-containing material. Potential starting materials include deciduous trees (such as birch, beech, poplar, alder and the like) and such plants or plant components as corn or maize, oat hulls, corn cobs and stalks, nut shells, straws, bagasse and cotton seed hulls. When wood is utilized as the starting material, it is preferably ground into chips, sawdust, shavings and the like and subjected to hydrolysis, or steam explosion and hydrolysis which creates hemicellulose material which can be used with the instant invention.

In addition to the above-listed materials, xylan or xylose-rich byproducts from wood processing procedures are also effective raw materials. For example, the spent liquor produced as a waste product from the production of wood pulp via the sulfite process - known as "sulphite-spent liquors" - contains undissolved wood solids, lignins, hexoses and pentoses, including xylose, and is an effective starting material for the production of xylitol. Other byproducts or waste products from paper or pulp processing which are xylan or xylose-rich can also be used.

In order to utilize the methods of the instant invention, an aqueous solution containing free xylose is required. Therefore, the hydrolysis of the starting

material by acids or enzymes to break down xylan to xylose may be required. For example, U.S. Patent Nos. 3,784,408 (Jaffe et al.) and U.S. Patent No. 3,586,537 (Steiner et al.) disclose methods for hydrolyzing xylan-containing material to produce xylose-rich solutions.

2. Fermentation of Xylose-containing aqueous solutions

The starting material may be treated before fermentation, if necessary, to remove any constituents which might be toxic or otherwise harm the yeast for fermentation. Whether such pretreatment is necessary depends on the starting material involved and the yeast which will be utilized for fermentation. Suitable pretreatment of raw material may include post-hydrolysis and/or separation steps. The concentration of xylose in the aqueous solution suitable for fermentation depends on the starting material utilized, but preferably is in the range of about 50 g/L to about 300 g/L.

To effect fermentation, the present invention utilizes a yeast strain which has the ability to convert xylose to xylitol, and most hexoses present to ethanol. Ethanol can be easily recovered by evaporation, distillation, or other known expedients much more simply and more efficiently than separation of xylose and/or xylitol from other sugars.

An example of a yeast strain which is suitable in this context is a *Candida tropicalis* deposited with the American Type Culture Collection (No. 9968). Other yeast strains include those of the genera *Candida*, *Pichia*, *Pachysolen* and *Debaryomyces*, (See N.J.W. Kregg-van Rij, *The Yeast. A Taxonomic Study*, 3 ed., Elsevier Science Publishers B.V. 1984.) which are capable of converting xylose to xylitol and hexoses to ethanol.

Prior to fermentation of the xylose-rich solution, the solution can be subjected to chromatographic separ-



ation to separate and remove larger molecules and ionized substances, when low purity liquids are used as raw material. For example, prefermentation chromatographic separation may be advantageous when sulfite spent liquors are used for the starting material.

Fermentation of the xylose-rich solution can take place in most commercially available batch fermenters equipped with aeration, agitation and pH control is preferred; for example, a Braun-Biostat [Model #E]. The preferred temperature for fermentation is between about 20°C and about 40°C, with a temperature of about 30°C being particularly preferred. Yeast cells are added to the xylose-rich solution; generally, the higher the concentration of yeast, the faster the fermentation will proceed. The optimum concentrations of yeast depends on the xylose liquor and its characteristics and xylose concentration in the liquor. We have found that adding yeast cells to a concentration of between about 0.1 g and about 10 g dry yeast/l (dry weight) substrate is preferred when the xylose concentration is between about 50 g/L and about 300 g/L.

Fermentation is enhanced by the addition of nutrients, and continues until most of the xylose has been converted to xylitol, and substantially all of the hexoses have been converted to ethanol; typically, fermentation will take between about 24 and about 144 hours, with a fermentation time of about 24-72 hours being particularly preferred. Using the method of the present invention, it is possible to convert over 90% of the xylose to xylitol.

### 3. Post-fermentation purification and xylitol separation

Following fermentation, the solution is clarified prior to separation of xylitol. In a batch fermentation process, the yeast cells are removed following the completion of fermentation. The removal of yeast cells can

be accomplished by centrifugation, filtration or similar expedients. Once the yeast cells have been removed and the solution is clear, the ethanol produced by fermentation can be removed by evaporation, distillation or other expedients at this stage.

After removal of the yeast cells (and possibly ethanol), the xylitol in the fermented solution is enriched by means of chromatographic separation. Such chromatographic separation is preferably carried out in a column packed with sulfonated polystyrene resin cross-linked with divinyl benzenes in an alkali/alkaline earth metal form. A method for large scale chromatography suitable for use in this context is described in U.S. Patent No. 3,928,193 (Melaja et al.). Chromatographic separation can also take place as a continuous process, utilizing a simulated moving bed process, as disclosed in U.S. Patent No. 2,985,589, also utilizing a DVB cross-linked sulfonated polystyrene resin.

Xylitol from xylitol-rich fractions derived from chromatographic separation can subsequently be crystallized in high yield utilizing conventional crystallization techniques including cooling and boiling crystallization. If cooling crystallization is used, the xylitol-rich fraction is seeded with xylitol crystals having an average diameter of about 30 microns, and the temperature of the solution is gradually lowered. The resulting crystals, preferably with an average diameter of about 250 to about 600 microns, are separated by centrifugation and washed with water to recover substantially pure crystalline xylitol.

B. Experimental

EXAMPLE I: Production of Xylitol from "Sulfite-Spent Liquor"

Xylitol was produced from a "sulfite-spent liquor" from hardwood utilizing fermentation with a *Debaryomyces*

hansenii yeast established as a strain which could ferment xylose to xylitol and most hexoses to ethanol. A batch of sulfite spent liquor (from the production of birchwood pulp) was treated as described in U.S. Patent No. 4,631,129 to obtain a xylose-rich fraction. The analysis of this fraction was as follows (carbohydrate composition measured by gas liquid chromatography analysis):

	Dry substance	30.4	w/w
10	pH	2.5	
	Calcium (CA ++)	2.0	% on dry substance (hereinafter "d.s.")
	Sodium (Na +)	0.1	% on d.s.
	Carbohydrates:		
15	Xylose	39.3	% on d.s.
	Arabinose	1.0	% on d.s.
	Rhamnose	1.2	% on d.s.
	Glucose	2.5	% on d.s.
	Mannose	0.1	% on d.s.
20	Galactose	2.0	% on d.s.

The fraction was neutralized with calcium oxide to a pH of about 6.2 by adding 10 g CaO per liter of the fraction. The fraction was then diluted to a concentration of 51 g xylose per liter of solution and then fermented with yeast cells. The fermentation was carried out in "shake" flasks (200 ml) at about 25°C for about 48 hours. The amount of yeast cells added was about  $1.7 \times 10^8$  cells per milliliter of fraction which were adapted to the fermentation solution after initial growth in xylose-rich solution. The yeast cells were removed after 48 hours by centrifugation and the resulting clear solution was subjected to chromatographic separation to separate xylitol produced by the fermentation under the following conditions:

## Composition of solution:

	Dry substance	24.0	w/w
	pH	5.9	
5	Carbohydrates:		
	Xylitol	24.7	% on d.s.
	Xylose	0.3	% on d.s.
	Glucose	2.1	% on d.s.
	Arabinose	0.6	% on d.s.
10	Calcium acetate	2.5	% on d.s.

## Column:

	Diameter	10	cm
	Height	200	cm
15	Resin	Zerolit 225 polystyrene-dvb-cation exchanger in calcium form, mean par- ticle size 0.32 mm, divinylbenze ("DVB") content 3.5%	
	Flow rate	50	ml/min
20	Temperature	65°C	
	Feed volume	500	ml

The elution profile from the column is shown in Figure 1. Samples were taken from the outflow and analyzed for dry substance and composition as set forth below in Table I with the outflow divided into three fractions:

TABLE I			
	Fraction	Fraction	Fraction
30	#1	#2	#3
Dry substance (grams)	134	10	58
Concentration (g/l)	12	10	16
Xylitol (% on d.s.)	0.7	40.0	79.0

35 Crystallization of xylitol produced by the fermentation method described above was accomplished as

follows. A xylitol-rich fraction was prepared as described above. The composition of a xylitol fraction utilized was:

	Dry substance	20 g per liter
5	Xylitol	86.6 % on d.s.
	Others	13.4 % on d.s.

From this solution xylitol was recovered by cooling crystallization. The solution was first evaporated to 86.5% dry substance concentration and transferred to a  
10 crystallizer equipped with cooling system and agitator. The initial temperature was about 65°C and the pH was about 5.3. The solution was seeded with xylitol crystals suspended in isopropanol with a crystal diameter of about 30 microns. The temperature was lowered during 3 hours  
15 from about 65°C to about 50°C. Under these conditions, the xylitol crystals grew to an average diameter of 250 micron. The crystals were separated from the solution by centrifugation and washed with water. The recovered crystals consisted of over 99% pure xylitol.

20

#### EXAMPLE 2: Production of Xylitol from Steam Exploded Birchwood

The raw material for this example was a steam-exploded birch wood hydrolyzate subjected to post-hydrolysis in order to break down the xylan to free xylose. For parameters of hydrolysis, refer to "Enzymatic hydrolysis of steam-pretreated lignocellulosic material,"  
25 Poutanen, K. and Puls, J., Proc. 3rd Eur. Congr. Biotechnol., Munich 1984, Vol. II, pp. 217-222. The composition  
30 of the resulting xylose-rich solution was following:

	Components	Concentration (g/l)
	Xylose	76
	Glucose	3.6
	Rhamnose	1.3
5	Mannose	2.1
	Galactose	2.4
	Arabinose	0.8

Total dry material was 15% by weight of the solution

The solution was fermented with *Candida tropicalis* yeast (ATCC 9968). The pH of the solution was adjusted to about 6 by the addition of 25% NaOH, and inoculated with 3 g/l yeast extract; 3 g/l malt extract and 5 g/l peptone were added as nutrients. The inoculum was prepared by growing the yeast in a 5% xylose solution with the same nutrient addition. During the fermentation the temperature was about 30°C. The fermentation was carried out in a Braun Biostat fermenter [Model #E] supplied with aeration (0.18 l/min) and agitation (200 rpm) and pH-control (25% NaOH) to maintain a pH of 6; the fermenter had a working volume of 8 l and a total volume 10 l. Foaming was controlled with Mazu 6000 antifoam agent. The analysis of samples from the fermentor is shown in Table II. Composition of samples from fermentation was analyzed by high performance liquid chromatography.

TABLE II

Time/hours		Yeast g/l	Xylose g/l	Xylitol g/l	Ethanol g/l
30	0	1.2	75.8	0.0	0.0
	24	1.4	66.5	4.8	3.7
	48	2.0	55.2	14.1	6.9
	72	2.6	37.3	29.4	7.7
	96	2.8	18.7	54.2	7.6
35	120	(not measured)	9.4	61.9	8.5
	144	(not measured)	3.0	52.7	6.7

Xylitol contained in the solution obtained by fermentation was concentrated in xylitol-rich fractions by chromatographic separation and crystallized to obtain 99% pure xylitol as in Example 1.

5

**EXAMPLE 3: Production of Xylitol from Steam Exploded Birchwood**

In Example 3, the raw material utilized was a steam exploded, posthydrolysate from birch wood according to the parameters discussed in Example 2. The composition of the solution was:

	Component	Concentration (g/l)
	Xylose	110.0
	Glucose	3.1
15	Rhamnose	3.5
	Mannose	3.4
	Galactose	1.5
	Arabinose	1.6

Total d.s. concentration was 15% by weight

20

Prior to fermentation the solution was subjected to a chromatographic separation process to remove most of large molecules and ionized substances. The chromatographic separation was carried out in column filled with AMBERLITE BH-1 (polystyrene-divinylbenzene) resin in sodium form. The conditions were as follows:

25

Resin: AMBERLITE BH-1 sulphonated polystyrene-divinylbenzene cross-linked with 5.5 % divb in sodium form.

Particle size 0.40 mm.

Column diameter 0.225 m and height 5.0 m.

30

Temperature: 65°C

Flow Rate: 0.04 m<sup>3</sup>/h

Feed: 18 l of a solution concentrated to 31.2 weight %.

35

The result of the chromatographic separation described above is shown graphically in Figure 2. Samples were taken at 5 minute intervals. The total cycle time

was 170 minutes. Composition of the feed solution into the separation column was as follows:

	Arabinose	0.6 % on d.s.
	Rhamnose	0.5 % on d.s.
5	Xylose	37.8 % on d.s.
	Mannose	1.2 % on d.s.
	Galactose	1.6 % on d.s.
	Glucose	1.6 % on d.s.
	Others	57.3 % on d.s.

10

The eluent was divided in 5 fractions. Fraction 2 was discarded from the process and fraction 4 was collected for the fermentation step. The remaining fractions were returned to the feed solution for the chromatographic separation to increase separation yield. The composition of the fractions is shown below in Table III.

15

Table III

Fraction	1	2	3	4	5
20 Time (minutes)					
(cut point)	15	100	115	140	155
Arabinose	1.3	0.1	0.2	1.0	2.1
Rhamnose	0.0	0.0	1.2	0.9	0.0
25 Xylose	24.8	3.4	57.6	71.2	46.6
Mannose	0.8	0.1	1.9	2.2	1.6
Galactose	0.8	0.1	2.0	2.0	1.4
Glucose	0.5	0.5	4.7	1.9	0.7
Other	71.9	95.8	32.3	20.3	47.6
30 (Numbers are expressed in terms of % on d.s.)					

30

The fermentation was carried out with *Candida tropicalis* yeast cells as in Example 2. The result of the fermentation is shown graphically in Figure 3. The obtained yield of xylitol was over 90 g/l from 100 g/l xylose. The nutrient used in this experiment was (Gistex)

35



yeast extract 15 g/l. The inoculum was grown in a hydrolyzate diluted with water (1/10) with 3% added glucose and 3% added yeast extract.

5 From the xylitol-rich solution the xylitol was recovered by chromatographic separation using a pilot scale column with the following characteristics:

Column: Height 4.5 m, diameter 0.225 m

Resin: Sulphonated polystyrene polymer cross-linked  
with 5.5% divinylbenzene. Mean particle size  
10 0.37 mm in Sodium form

Flow rate: 0.03 m<sup>3</sup>/h

Temperature: 65°C

Feed Solution: 24 g kg of a 24 weight % solution (dry substance 5.76 kg).

15 Composition:

Xylitol 64.0 % on d.s.

Xylose 2.0 % on d.s.

Arabinose

+ mannose 1.4 % on d.s.

20 Galactose

+ rhamnose 1.2 % on d.s.

Glucose 0.4 % on d.s.

Mannitol 0.9 % on d.s.

Other 30.1 % on d.s.

25 The separation is presented graphically in Fig. 4. Five fractions were recovered. The compositions are shown below in Table IV. Fraction No. 4 was the product fraction from which pure xylitol was crystallized as set forth in Example 1.

Table IV

Fraction	1	2	3	4	5
Time min					
(cut point)	10	100	105	155	165
5 Xylitol	8.7	4.9	56.3	85.0	55.6
Xylose	0.2	2.0	4.4	2.0	0.4
Arabinose					
+ mannose	3.6	0.5	1.4	1.7	4.2
10 Galactose					
+ rhamnose	0.3	0.6	3.8	1.4	0.5
Glucose	0.0	0.1	1.9	0.4	0.0
Mannitol	0.2	0.2	2.0	1.1	0.7
Other	87.1	81.8	30.2	8.5	38.5
15 (Numbers are given as % on d.s.)					

#### EXAMPLE 4: Crystallization of Xylitol from Fermented Solution

Xylitol was crystallized from a xylitol-rich solution recovered by chromatographic separation from the fermented solution. The solution which contained 82.5% xylitol of the dry substance was evaporated at 65°C temperature to 92% concentration. 2200 g of the evaporated solution was seeded with 0.04 mm xylitol seed crystals. The seed amount was 0.03%. The temperature of the solution was lowered to 45°C in 55 hours according to a pre-determined program:

$$T = T_1 - (t : t^1)^2 \times (T_1 - T_2)$$

where

30 T = the temperature of the solution C

T<sub>1</sub> = the temperature at seeding (65°C)

T<sub>2</sub> = the final temperature (45°C)

t = time from the seeding in hours

t<sub>1</sub> = total time of crystallization (55 h)

5 The crystallization was carried out in a vertical crystallizer equipped with a mixer. The crystals were separated from the solution by centrifugation (5 minutes, 2000 g) and washed with water. The recovered crystals had a median size of 0.37 mm and a purity of 99.4% (HPLC).

10 The foregoing general discussion and experimental examples are intended to be illustrative of the present invention, and are not to be considered as limiting. Other variations within the spirit and scope of this invention are possible, and will present themselves to those skilled in the art.

## WE CLAIM:

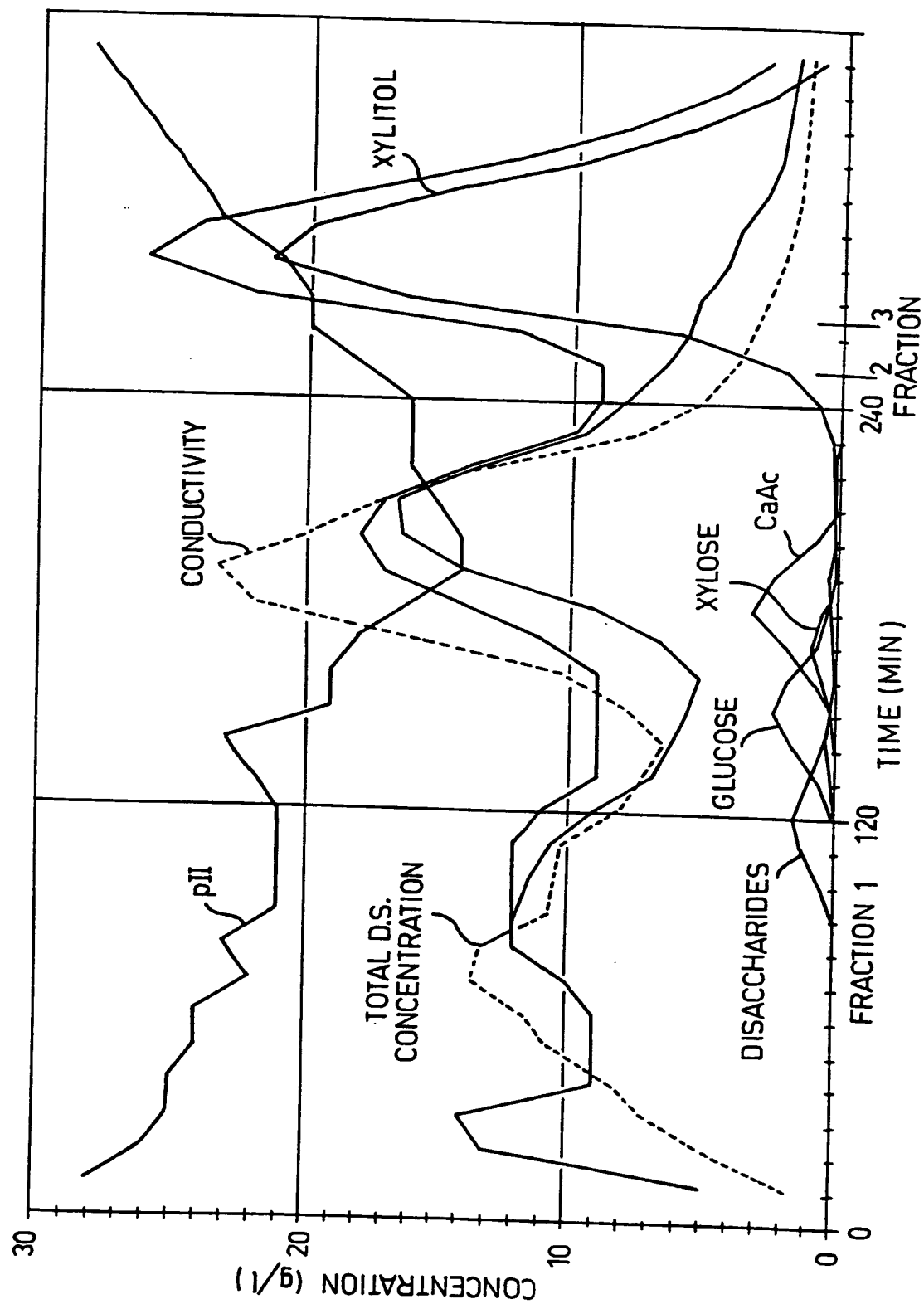
1. A method for the production of xylitol from an aqueous xylose solution which comprises the steps of:
  - 5 fermenting said aqueous xylose solution with a yeast strain capable of converting free xylose to xylitol and free hexoses present in said solution to ethanol for a period of time sufficient to produce a fermented solution containing xylitol; and
  - 10 separating xylitol-rich fraction or fractions by chromatographic separation from said fermented solution.
2. The method of claim 1 wherein said yeast genera is *Candida* or *Debaryomyces*.
3. The method of claim 2 wherein said yeast strain is  
15 *Candida tropicalis*.
4. The method of claim 3 wherein said yeast strain is *Candida tropicalis* having American Type Culture Collection Accession Number 9968.
5. The method of claim 2 wherein said yeast strain is  
20 *Debaryomyces hansenii*.
6. The method of claim 1 wherein xylitol is recovered from said xylitol rich fraction or fractions.
7. The method of claim 6 wherein said xylitol is recovered from said xylitol rich fraction or fractions by  
25 crystallization.
8. The method of claim 1 wherein said fermentation is carried out at about 30°C. for about 48 hours.
9. The method of claim 7 wherein said fermentation is carried out for about 48 hours.
- 30 10. The method of claim 1 wherein said yeast cells are removed by centrifugation after fermentation and before chromatographic separation.
11. The method of claim 1 wherein said yeast cells are removed by filtration after fermentation and before  
35 chromatographic separation.

12. The method of claim 1 wherein said chromatographic separation utilizes a cation-exchange resin as column filling material.
13. The method of claim 12 wherein the resin has a divinyl benzene cross-linked sulfonated polystyrene skeleton.
14. The method of claim 1 wherein water is used as eluent in chromatographic separation.
15. The method in accordance with claim 1 wherein said aqueous xylose solution is a hemicellulose hydrolysate.
16. The method of claim 1 wherein said aqueous xylose solution is a xylose enriched fraction from sulfide spent liquor.
17. The method in accordance with claim 1 wherein ethanol is removed by evaporation or distillation after fermentation.
18. The method of claim 1 wherein said fermentation is carried out in medium where xylose concentration is ranging from 50 g/l to 300 g/l.
19. The method of claim 1 wherein said fermentation is carried out at a pH ranging from 3 to 9.
20. The method of claim 19 wherein said fermentation is carried out at a pH ranging from about 5 to 7.
21. The method of claim 1 wherein said fermentation is carried out at a temperature ranging from about 10°C to about 45°C.
22. The method of claim 21 wherein said fermentation is carried out at a temperature ranging from about 25°C to 35°C.
23. The method of claim 1 wherein said fermentation is carried out with limited oxygen supply.
24. The method of claim 1 wherein components which may be toxic to the yeast used for fermentation or other impurities are separated from said aqueous xylose solution prior to fermentation.
25. The method of claim 24 wherein said separation is carried out by chromatographic separation means.



FIG. 1

## ELUTION PROFILE FOR FERMENTED HYDROLYZATE

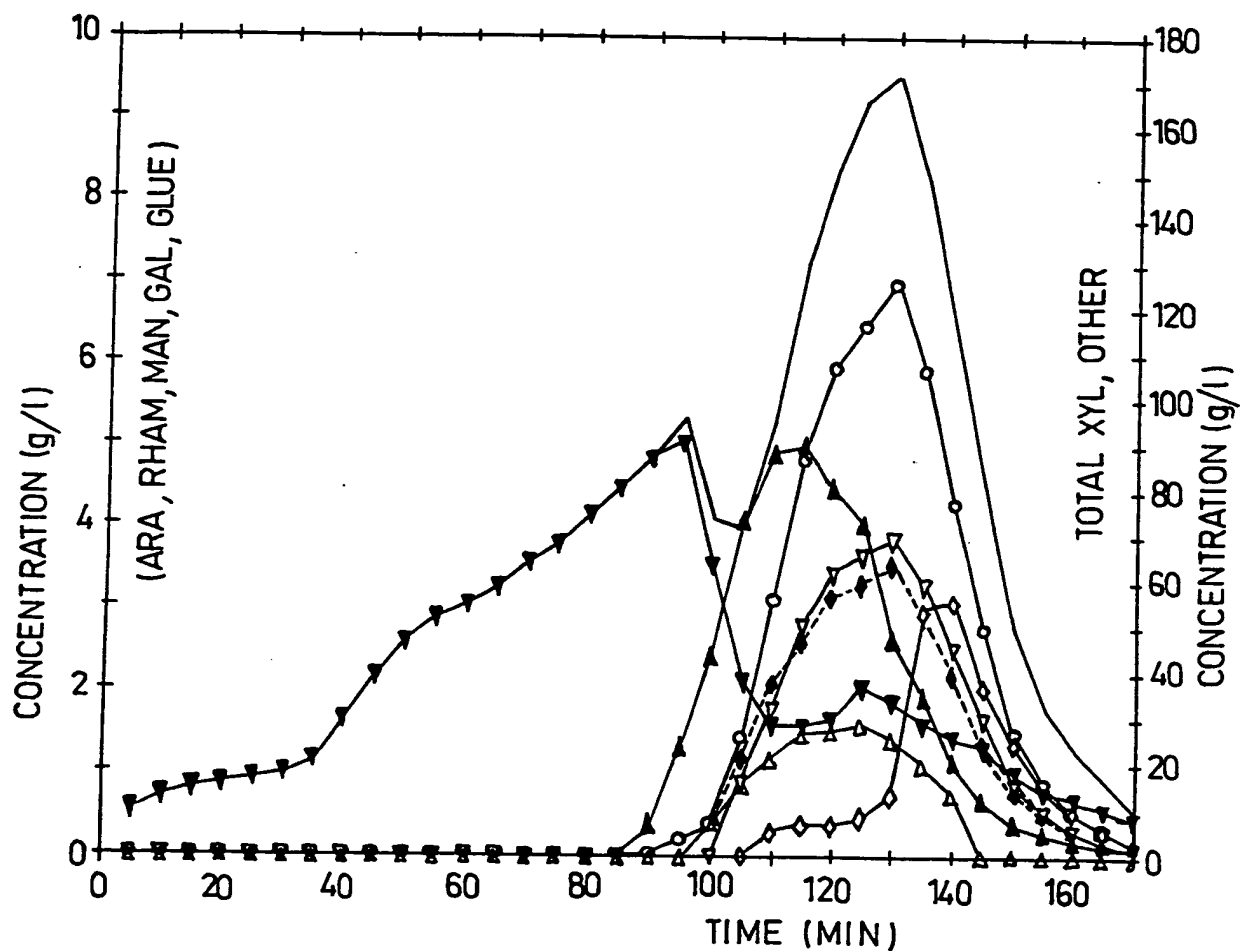


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FIG. 2

ELUTION PROFILE FOR SEPARATION  
BY COLUMN CHROMATOGRAPHY

KEY:

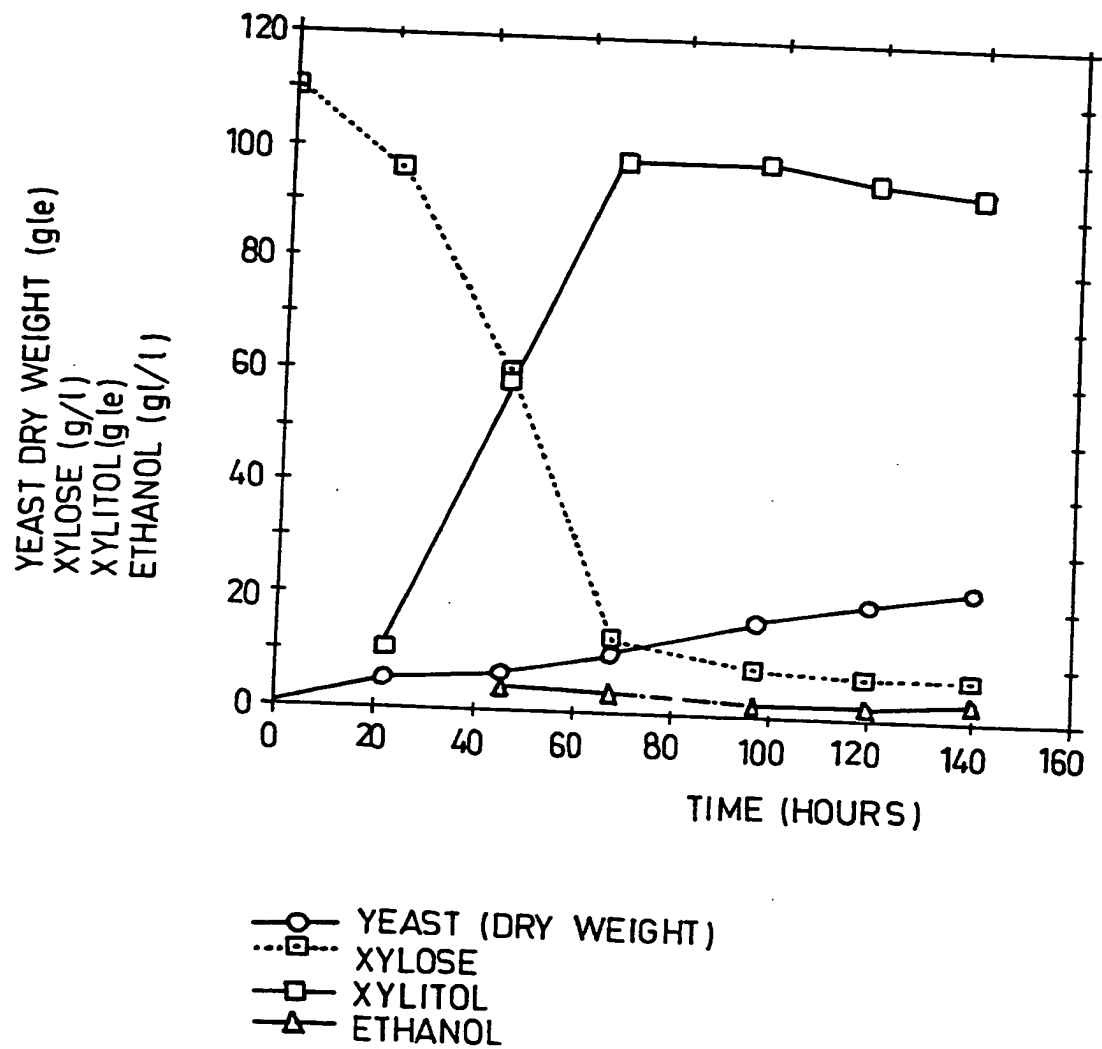
— TOTAL  
—○— XYLITOL  
—◇— ARA  
—△— RHAM  
—▽— MAN

—◆— GAL  
—▲— GLUE  
—▼— OTHER

SUBSTITUTE SHEET



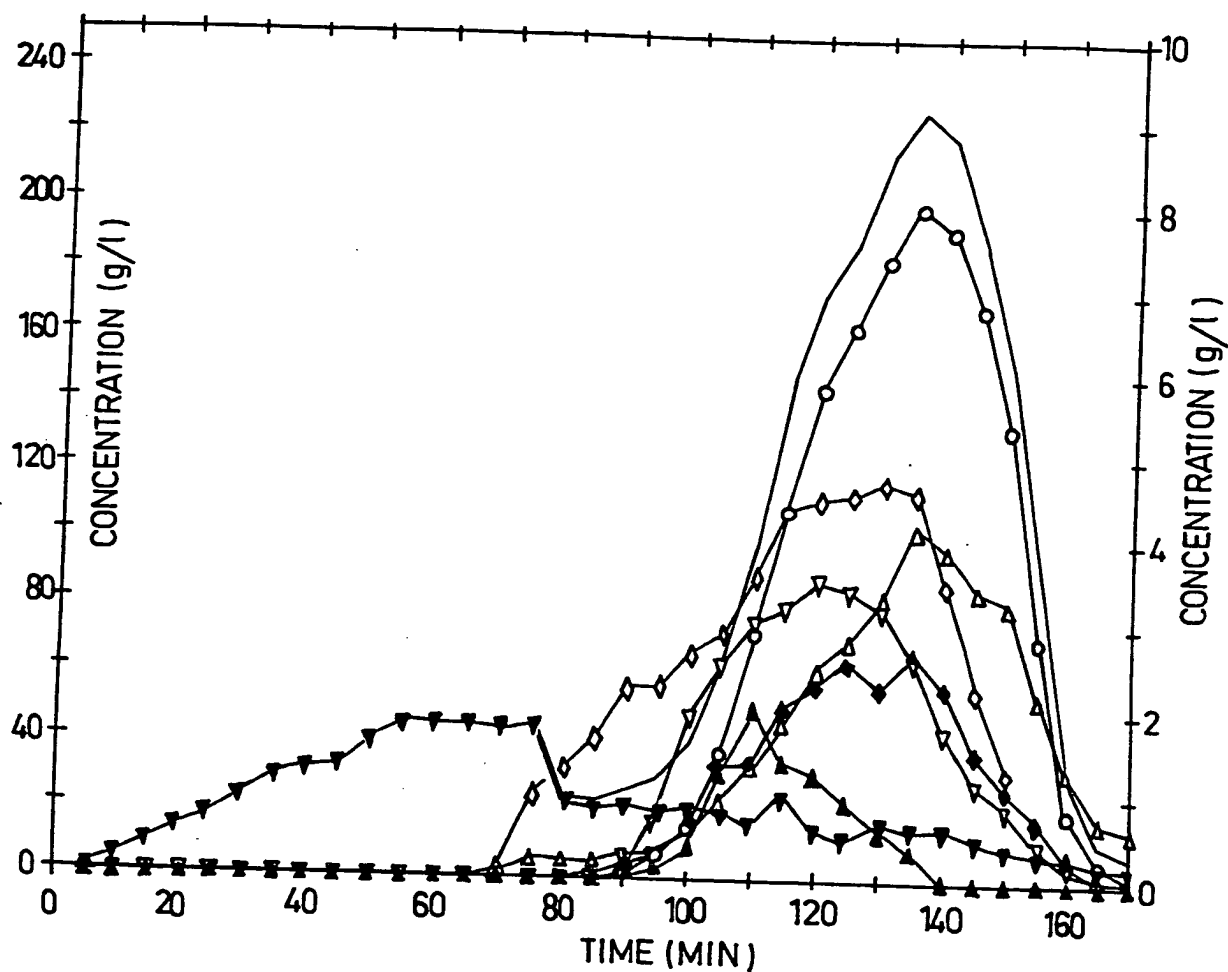
FIG. 3

FERMENTATION OF XYLOSE SOLUTION  
OF CANDIDA TROPICALIS

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FIG. 4

ELUTION PROFILE FOR SEPARATION  
BY COLUMN CHROMATOGRAPHY

KEY:

— TOTAL  
—○— XYLITOL  
—◇— XYLOSE

—△— ARA +  
MANNOSE  
—▽— GALA +  
RAM

—▲— GLU  
—◆— MANNITOL  
—▼— OTHER

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# INTERNATIONAL SEARCH REPORT

International Application No PCT/FI 90/00015

<b>I. CLASSIFICATION OF SUBJECT MATTER</b> (if several classification symbols apply, indicate all) <sup>6</sup> According to International Patent Classification (IPC) or to both National Classification and IPC IPC5: C 12 P 19/02, C 12 P 7/18		
<b>II. FIELDS SEARCHED</b>		
Minimum Documentation Searched <sup>7</sup>		
Classification System	Classification Symbols	
IPC5	C 12 P	
Documentation Searched other than Minimum Documentation to the Extent that such Documents are Included in Fields Searched <sup>8</sup>		
SE,DK,FI,NO classes as above		
<b>III. DOCUMENTS CONSIDERED TO BE RELEVANT<sup>9</sup></b>		
Category *	Citation of Document, <sup>11</sup> with indication, where appropriate, of the relevant passages <sup>12</sup>	Relevant to Claim No. <sup>13</sup>
X	Dialog Information Services, File 351, World Patent Index 81-90, Dialog accession no. 88-297740/42, Horitsu H: "Sugar alcohol prepn. - by treating mixed sugar soln. contg. substrate sugar and hydrogen donor sugar with Candida yeast", JP 63219386, A, 880913, 8842, (Basic) --	1-25
X	Agr.Biol.Chem., Vol. 30, No. 11, 1966 Hiroshi Onishi et al: "The Production of Xylitol. L-Arabinitol and Ribitol by Yeasts ", see page 1139 - page 1144 --	1-25
P,X	Chemical Abstracts, volume 112, no. 5, 29 January 1990, (Columbus, Ohio, US), Amaral-Collaco, M.T. et al : "Utilization of the hemicellulosic fraction of agro industrial residues by yeasts ", see page 449, abstract 34371t, & Enzyme Syst.Lignocellul.Degrad. 1989,, 221- 230 --	1-25
* Special categories of cited documents: <sup>10</sup> <div style="display: flex; justify-content: space-between;"> <div style="width: 45%;"> <p>"A" document defining the general state of the art which is not considered to be of particular relevance</p> <p>"E" earlier document but published on or after the international filing date</p> <p>"L" document which may throw doubts on priority claim(s) or which is cited to establish the publication date of another citation or other special reason (as specified)</p> <p>"O" document referring to an oral disclosure, use, exhibition or other means</p> <p>"P" document published prior to the international filing date but later than the priority date claimed</p> </div> <div style="width: 45%;"> <p>"T" later document published after the international filing date or priority date and not in conflict with the application but cited to understand the principle or theory underlying the invention</p> <p>"X" document of particular relevance, the claimed invention cannot be considered novel or cannot be considered to involve an inventive step</p> <p>"Y" document of particular relevance, the claimed invention cannot be considered to involve an inventive step when the document is combined with one or more other such documents, such combination being obvious to a person skilled in the art</p> <p>"&amp;" document member of the same patent family</p> </div> </div>		
<b>IV. CERTIFICATION</b>		
Date of the Actual Completion of the International Search	Date of Mailing of this International Search Report	
25th April 1990	1990 -04 27	
International Searching Authority	Signature of Authorized Officer	
SWEDISH PATENT OFFICE	<i>Yvonne Siösteen</i> Yvonne Siösteen	

III. DOCUMENTS CONSIDERED TO BE RELEVANT (CONTINUED FROM THE SECOND SHEET)		
Category *	Citation of Document, with indication, where appropriate, of the relevant passages	Relevant to Claim No
X	Chemical Abstracts, volume 109, no. 23, 5 December 1988, (Columbus, Ohio, US), see page 545, abstract 209692r, & FI,, 76377 (Farmos-Yhtymä Oy) 1988 --	1-25
X	Chemical Abstracts, volume 94, no. 25, 22 June 1981, (Columbus, Ohio, US), Gong, Cheng-Shung et al : "Quantitative production of xylitol from D-xylose by a high-xylitol producing yeast mutant <i>Candida tropicalis</i> HXP2 ", see page 462, abstract 207106a, & Biotechnol.Lett. 1981, 3(3), 125- 130 --	1-25
X	Chemical Abstracts, volume 102, no. 4, 28 January 1985, (Columbus, Ohio, US), Ermakova, I.T. et al : "Characteristics of biosynthesis of polyols by the yeast <i>Candida guilliermondii</i> grown in media with different carbon sources ", see page 290, abstract 3048g, & Mikrobiologiya 1984, 53(5), 803- 808 --	1-25
A	WO, A1, 88/05467 (FARMOS-YHTYMÄ OY) 28 July 1988, see the whole document -- -----	1-25



ANNEX TO THE INTERNATIONAL SEARCH REPORT  
ON INTERNATIONAL PATENT APPLICATION NO. PCT/FI 90/00015

This annex lists the patent family members relating to the patent documents cited in the above-mentioned international search report. The members are as contained in the Swedish Patent Office EDP file on **90-03-30**. The Swedish Patent Office is in no way liable for these particulars which are merely given for the purpose of information.

Patent document cited in search report	Publication date	Patent family member(s)	Publication date
WO-A1- 88/05467	88-07-28	NONE	

